Biomedical Optics EXPRESS

Deep learning-based adaptive optics for light sheet fluorescence microscopy: supplement

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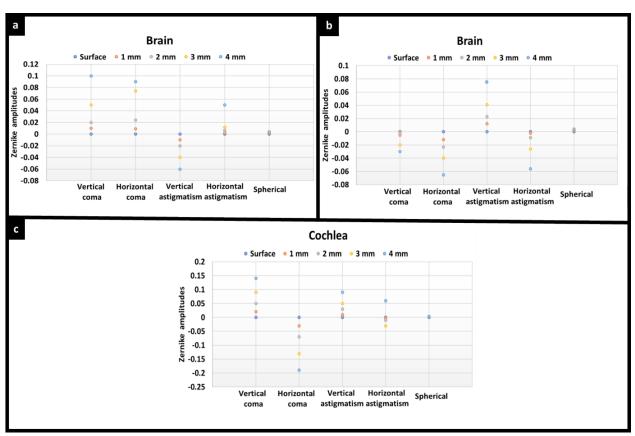
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Supplement DOI: https://doi.org/10.6084/m9.figshare.22954616

Parent Article DOI: https://doi.org/10.1364/BOE.488995

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Supplementary information

Fig. S1. Depth-dependent Zernike amplitudes for (a-b) two rat brains and (c) pig cochlea. Please note that the values provided are highly dependent on the quality of the clearing and age of the sample. Therefore, these values may vary from sample to sample, as well as within a sample depending on the spatial location that is imaged.

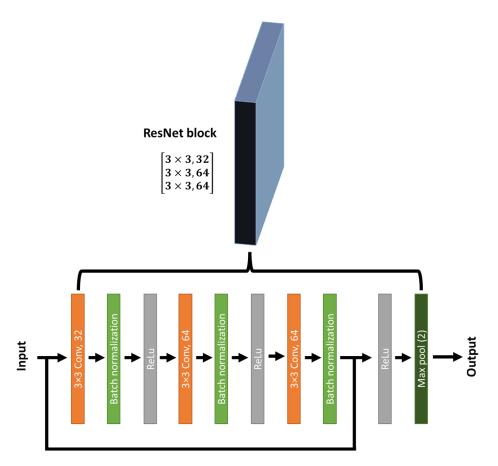


Fig. S2. The architecture of a single ResNet block.

Zernike Amplitudes							STD
Z ₄ (Vertical Coma)	-0.0590	-0.0572	-0.0545	-0.0504	0.0505	-0.0543	0.0040
Z₅ (Horizontal Coma)	0.0216	0.0297	0.0296	0.0249	0.0280	0.0267	0.0035
Z ₆ (Vertical Astigmatism)	0.0414	0.0442	0.0492	0.0479	0.0491	0.0464	0.0034
Z ₇ (Horizontal Astigmatism)	-0.0058	-0.0071	-0.0108	-0.0045	-0.0137	-0.0084	0.0038
Z ₁₀ (Spherical)	0.0114	0.0113	0.0114	0.0107	0.0105	0.0111	0.0004

Fig. S3. The estimated Zernike amplitudes for different locations within the FOV (color coded), the mean and standard deviation (STD) are shown.

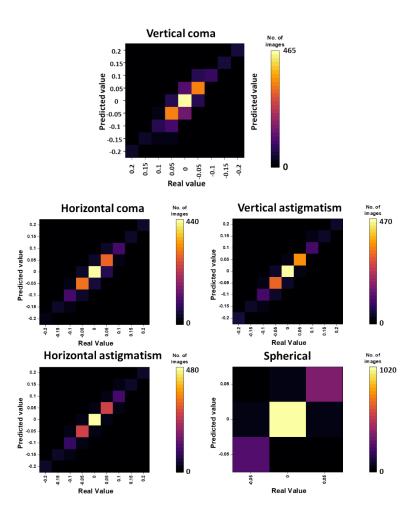


Fig. S4. Confusion matrices for different aberration types. The matrices were obtained from the *non-shared* network.

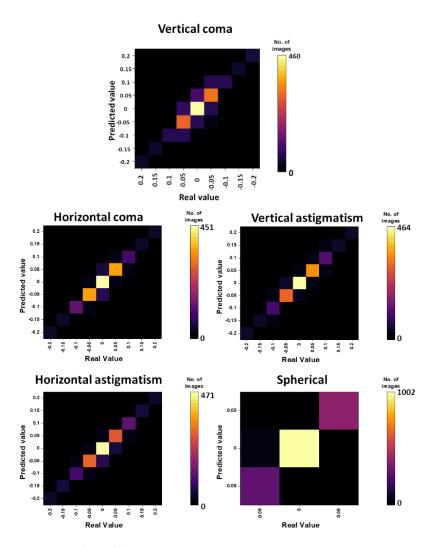


Fig. S5. Confusion matrices for different aberration types. The matrices were obtained from the *shared* network.

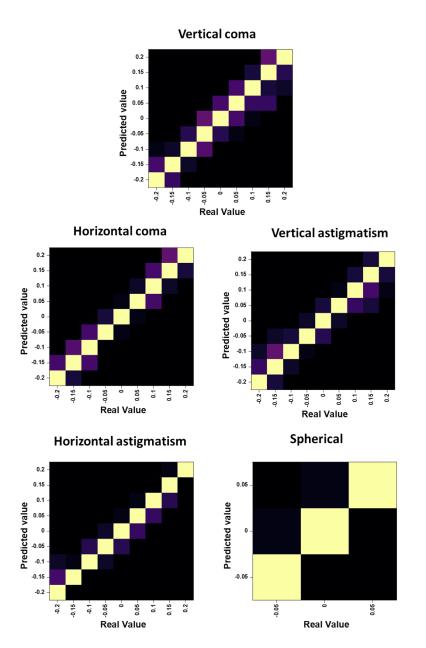
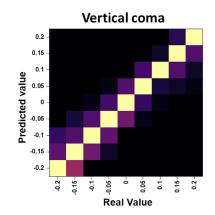


Fig. S6. Normalized confusion matrices for different aberration types. The matrices were obtained from the non-*shared* network.



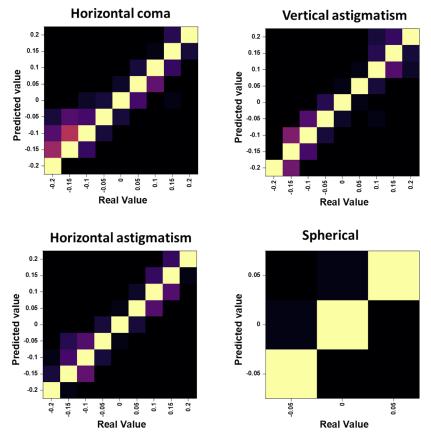


Fig. S7. Normalized confusion matrices for different aberration types. The matrices were obtained from the *shared* network.

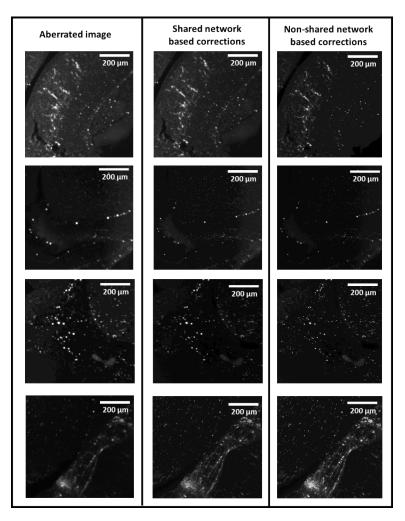


Fig. S8. Additional examples of aberration correction using the *non-shared* and *shared* networks. The figure shows data acquired from tissue cleared rat brains.